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ECURITY CLASSIFICATION OF THIS PAGE

REPORT DOCUMENTATION PAGE					Form Approved OMB No. 0704-0188			
1a. REPORT SECURITY CLASSIFI (U)	1b. RESTRICTIVE MARKINGS NA							
2a. SECURITY CLASSIFICATION NA	3. DISTRIBUTION/AVAILABILITY OF REPORT							
2b. DECLASSIFICATION / DOWN-NA	Distribution Unlimited							
4. PERFORMING ORGANIZATIO	5. MONITORING ORGANIZATION REPORT NUMBER(\$)							
Massachusetts Ins	NA							
6a. NAME OF PERFORMING OF Massachusetts Ins of Technology		6b. OFFICE SYMBOL (If applicable)	7a. NAME OF MONITORING ORGANIZATION Office of Naval Research					
6c. ADDRESS (City, State, and	7b. ADDRESS (City, State, and ZIP Code)							
77 Massachusett	800 North Quincy Street							
Cambridge, Massachusetts 02139			Arlington, Virginia 22217-5000					
a. NAME OF FUNDING / SPONSORING Bb. OFFICE SY			9. PROCUREMENT INSTRUMENT IDENTIFICATION NUMBER					
ORGANIZATION Office of Naval R	esearch	(If applicable) ONR	N00014-84-K-0300					
Bc. ADDRESS (City, State, and Z				10. SOURCE OF FUNDING NUMBERS ,				
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Arlington, Virginia 22217-5000			61153N	RR04106	441		Ì	
		tion in the Ce	ll Cycle					
12. PERSONAL AUTHOR(S) Alexander Rich								
13a. TYPE OF REPORT	13b. TIME C			14. DATE OF REPORT (Year, Month, Day) 15 PAGE COUNT			OUNT	
ANNUAL 16 SUPPLEMENTARY NOTATION		/87to_9/88_	10/14/8	38		9		
16 SUPPLEMENTARY NOTATIO	,							
17. COSATI CO	DES	18. SUBJECT TERMS (Continue on reverse if necessary and identify by block number)						
FIELD GROUP	SUB-GROUP	DNA CONFORMATIONAL CHANGES, PROPELLER (P) DNA, Z-DNA, HOMOLOGOUS RECOMBINATION						
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20. DISTRIBUTION/AVAILABILITY OF ABSTRACT		21. ABSTRACT SECURITY CLASSIFICA	TION
UNCLASSIFIED/UNLIMITED SAME AS RPT	DTIC USERS	(U)	
27a. NAME OF RESPONSIBLE INDIVIDUAL M. Marron		22b TELEPHONE (Include Area Code) (202) 696–4760	22c OFFICE SYMBOL ONR

DD Form 1473, JUN 86
DISTRIBUTION STATEMENT A

Previous editions are obsolete.

SECURITY CLASSIFICATION OF THIS PAGE

Approved for public release:

Distribution Unlimited

Report on Contract No. N00014-84-K-0300

Principal Investigator: Alexander Rich

Contractor: Massachusetts Institute of Technology

Contract Title: Transcriptional Regulation in the Cell Cycle

Start Date: 1 June 1987

Objective

We explore the role of conformational changes in DNA both in regulating transcription as well as influencing homologous genetic recombination. One of the important events associated with transcriptional activation is the formation of DNA loops in regulatory regions. Looping brings together regulatory proteins that are found at different positions along the regulatory segment of DNA. The looping involves bending of the DNA at specific segments. It has been known that segments of oligo(dA)·oligo(dT) are associated with DNA bending. Recently, we have solved the structure of a DNA dodecamer that incorporates a segment of d(A3T3)·d(T3A3). Knowledge of the details of this conformation suggest they may be important in providing bending points in DNA looping.

Z-DNA affinity columns have been used for isolating Z-DNA binding proteins. Recently we have found such proteins that are active in carrying out the strand transferase activity that is involved in homologous recombination. (f(x))

Progress Report

For some time it has been known that DNA is capable of adopting a bent conformation. The sequence oligo(dA)·oligo(dT) is frequently associated with DNA bending. This is pointed out most strikingly in the DNA minicircles that are found in trypanosome in which the segments of the five base pairs of dA5·dT5 are interspersed into the minicircle every ten base pairs. It is the presence of these segments that makes possible a circularization of DNA with a small number of nucleotides. DNA looping is found in the upstream regulatory regions of DNA. This looping is associated with activation of DNA for transcription. Looping phenomena bring together proteins attached to DNA that are widely separated when the DNA is linear. The looping of DNA is associated with activation of transcription.

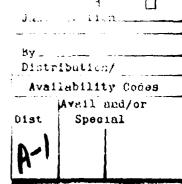
In addition, looping is frequently stabilized by oligo(dA)·oligo(dT) stretches that are found in upstream regulatory regions.

In order to study this problem we have solved the structure of a fragment of a DNA dodecamer that contains a d(A3T3)·d(T3A3) segment in it. Segments of DNA with only AT base pairs have a narrow minor groove. This groove can serve as a binding site for certain planar molecules. We have solved the structure of a DNA dodecamer d(CGCA3T3GCG). both by itself and bound to the antibiotic distamycin. The distamycin molecule was found bound in the minor groove of DNA. However, the A3T3 segments of the molecule had an unusual conformation both while the distamycin was present and in the DNA itself. In this conformation, the AT base pairs were found to have a strong propeller twist. This propeller twist was stabilized by bifurcated or three-centered hydrogen bonds between adenine amino NH2 and two thymine O4 atoms on the opposite strand (see Fig. 1). The propeller twist of the base pairs put the adenine amino group into a position where it could make two hydrogen bonds rather than one.

DNA bending is associated with oligo(dA)-oligo(dT) segments. This segment of DNA in the propeller conformation is likely to have a bend at the interface between the propeller conformation and the B-DNA outside of it. The reasons for this are associated with the important stacking interactions between the bases. In the three-dimensional structure of yeast tRNAPhe, the anticodon stem is at an angle compared to the adjacent dihydro U stem. Between these two stems is an AG base pair that is frozen in the propeller conformation. The anticodon stem stacks on one blade of the propeller, while the dihydro U stem stacks on the other blade of the propeller, thus giving rise to an angle in the orientation of these two adjacent helical segments of the molecule.

A similar situation may prevail in DNA with an oligo(dA)-oligo(dT) segment. The B-DNA segment with planar base pairs cannot stack upon both the adenine and the thymine bases in the propeller conformation. However, it may stack predominately on one blade of the propeller, which would have the effect of giving rise to an angle or bend in the helical axis of the B-DNA segment relative to the propeller DNA segment. A similar distortion could take place at the other end of the otherwise straight propeller segment. Thus, the propeller conformation may serve as an important component in stabilizing DNA looping.





Role of Z-DNA in Homologous Recombination

Homologous recombination is a process whereby two DNA molecules come together at a position where their nucleotide sequences are identical. The recombination process results in a "crossing over" of the duplex molecules. The mechanism of homologous recombination is not clearly understood. One mechanism that has been described involves an initial scission of a polynucleotide chain followed by unwinding of a DNA strand and invasion of a duplex DNA by this single strand. Such a process could ultimately lead to the formation of a holiday junction and recombination.

However, there is likely to be an alternative recombination pathway in which DNA strands sense the homology or nucleotide sequence identity by forming a recombination intermediate involving both right-handed and left-handed Z-DNA in this recombination intermediate. A process such as that, outlined in Figure 2, would involve the participation of proteins that have the ability to recognize Z-DNA. Several recombination enzymes have already been reported that have the ability to bind to Z-DNA.

We have looked for the presence of recombination proteins that bind to Z-DNA in human acute lymphoblastic leukemia cells. To do this we have constructed a Z-DNA affinity column as outlined in Figure 3. In this column the Z-DNA segments are held through biotin-Avidin linkers only at the ends of the Z-DNA segments. A column such as this was used to isolate proteins from the nuclei of human cells. The proteins were assayed for their ability to carry out strand transferase, that is assayed by measuring the combination of a single-stranded circular DNA molecule with a linearized duplex (Fig. 4). The strand transferase activity was detected in the proteins eluted from the Z-DNA affinity column. This strand transferase was characterized both in the electron microscope and in control experiments that identified its dependence on DNA sequence homology. experiments demonstrated the directionality of the reaction. on ATP was also found associated with this enzymatic activity. Even though the enzyme is not yet purified, the fraction containing the enzyme has considerable binding to Z-DNA as compared to B-DNA.

Objectives for the Next Year

1. Propeller DNA: We plan to extend the investigations on the propeller conformation of DNA by seeing whether the fiber diffraction data obtained from fibers of poly(dA)·poly(dT) are fully consistent with the propeller conformation that we have found in the DNA dodecamer crystals. This will be necessary in order to demonstrate the occurrence of the propeller conformation in this sequence. It will also be needed to explain the unusual physical properties of poly(dA)·(dT) compared to the

alternating copolymer poly(dAT). The poly(dA)-poly(dT) melts almost 8 degrees higher than poly(dAT). This additional stability is probably associated with the formation of bifurcated hydrogen bonds as well as other changes associated with the propeller conformation.

Attempts will also be made to look for other DNA conformational changes that are associated with transcriptional activation. Some of the candidates that are being looked at at the present involved homopurine-homopyrimidine sequences that are known to undergo conformational changes associated with transcriptional activation.

2. Z-DNA and Homologous Recombination: Our objective here is to continue the purification of the enzymatic activity that has been detected with a view toward understanding the manner in which this strand transferase activity participates in homologous recombination. Purification of the enzyme would also make it possible to understand which other enzymatic activities may be needed to carry out the recombination step. The other enzymatic activities involved include resolvases that are known to cleave the Holiday junctions and religate the severed polynucleotide chains to form the recombined molecule. The material isolated from the Z-DNA affinity column is likely to contain, in addition to Z-DNA binding proteins, other proteins that bind to Z-DNA binding proteins. Some of these may include enzymes such as resolvases as well as ligases that are important in the recombination process.

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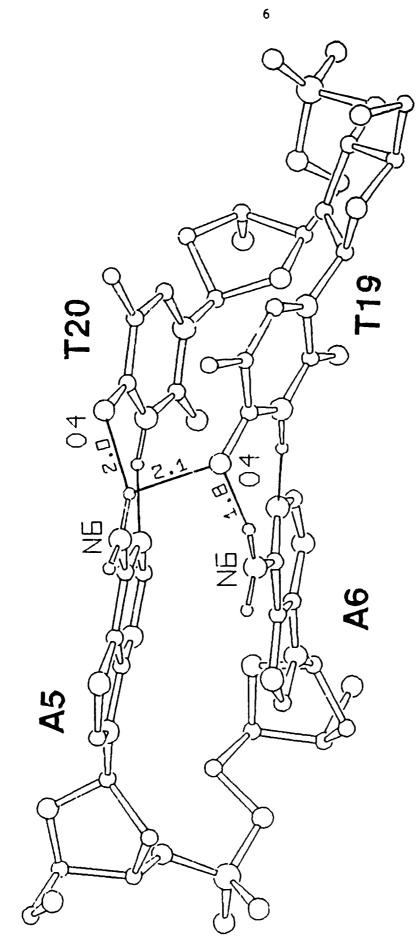
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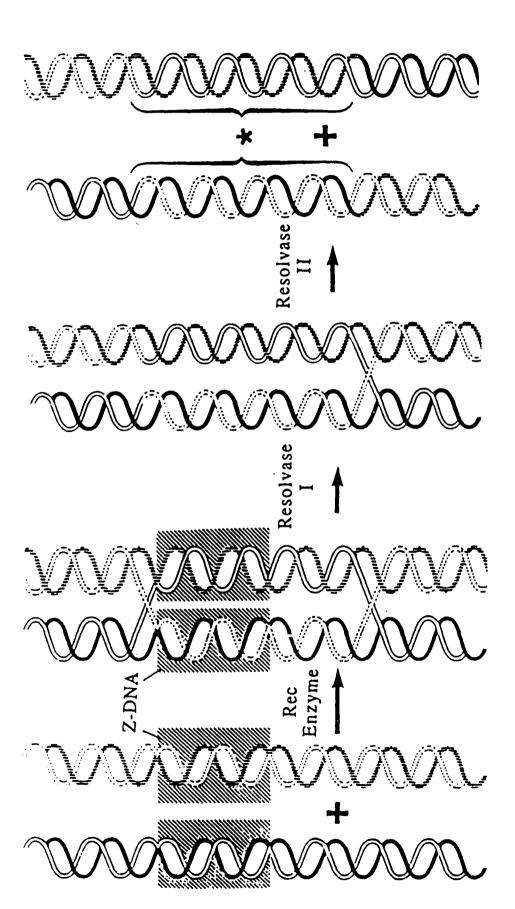
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Bifurcated Hydrogen Bond Lengths with Hydrogen Atoms

PARANEMIC PATHWAY IN HOMOLOGOUS RECOMBINATION



Recombined Molecules *Heteroduplex DNA

Holliday Junction

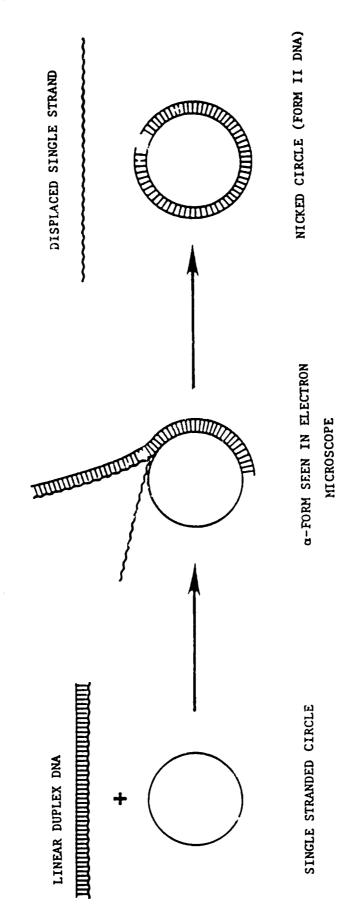
Recombinant

Identical Sequences Molecules with

Paranemic Joint

- A (Avidin) Biotin'-Z- DNA 4 Ø A: BX Preparation of Z-DNA Affinity Column Agarose – Steptavidin B (Biotin) ၁၁ Biotin-dCTP dGTP 4MNaCI Br2 Biotin Z-DNA Hinp Z-DNA B-DNA B-DNA poly (dG-dC) B-DNA polymerase Klenow ၁၁ 8

STRAND TRANSFERASE



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